

JPPIPA 8(2) (2022)

Jurnal Penelitian Pendidikan IPA

Journal of Research in Science Education



http://jppipa.unram.ac.id/index.php/jppipa/index

Estimation of Inheritance of Kenaf Resistance Against Root Chole Nematodes (*Meliodogyne incognita*)

Parnidi¹, Aditya Aji Noftara², Mala Murianingrum¹, Taufiq Hidayat RS¹, Marjani¹

¹ Sweetener and Fiber Crops Research Institute, Malang, Indonesian ² Agriculture Faculty - Brawijaya University, Malang, Indonesian

DOI: <u>10.29303/jppipa.v8i2.1211</u>

Article Info

Received: December 7, 2021 Revised: April 17, 2022 Accepted: April 19, 2022 Published: April 30, 2022 **Abstract:** Root puru nematodes (*Meloidogyne spp.*) on kenaf plants are known as one of the harmful plant-disturbing organisms. NPA infection in roots causes plants to grow stunted and production potential decreases. The inheritance of kenaf resistance to NPA *M. incognita* was carried out using a sixpopulation approach (P1, P2, F1, F2 BC1P1 and BC1P2). The genotype (Kenafindo Agribun 2 = Kin2) which was highly susceptible to NPA *M. incognita* was used as the female parent and the tolerant genotype (Karangploso 15 = KR15) as the male parent. The nature of kenaf resistance to NPA *M. incognita* was based on the variable number of shells, reproductive factors, number of 2nd instar juveniles, number of egg masses and number of eggs per egg mass. Based on the results of the study showed that the resistance of kenaf to NPA *M. incognita* was not influenced by female parents. The number of resistance to NPA *M. incognita* was additive-dominant gene. The action of genes controlling resistance to NPA *M. incognita* was additive-dominant. The heritability value in the broad sense and the heritability value in the narrow sense for all variables of kenaf resistance to NPA *M. incognita* were high.

Keywords: Heritability; Number of puru; Reproductive factors; Kenaf crosses.

Citation: Parnidi, P., Noftara, A.A., Murianingrum, M., Hidayat, R.T., & Marjani, M. (2022). Estimation of Inheritance of Kenaf Resistance Against Root Chole Nematodes (Meliodogyne incognita). *Jurnal Penelitian Pendidikan IPA*, 8(2), 691–698. <u>https://doi.org/10.29303/jppipa.v8i2.1211</u>

Introduction

Namatoda Puru Root (*Meloidogyne spp.*) in kenaf is known as one of the plant pest organisms (OPT) that causes plants to grow stunted, thereby reducing production potential. (Tahery et al., 2011; Wokoma et al., 2015; Adegbite, 2018). According to Lawrence & McLean (1992) the loss of yield of kenaf plants due to root-knot nematode infection reached 32% – 67%. Tahery et al. (2011) added that the population of root-knot nematodes of 5000/500 cm3 of soil could reduce plant height, stem diameter, stem weight and kenaf roots. As a result of root-knot nematode infection, the wet weight of kenaf stems can be reduced up to 3 tons per hectare. The population of forty juveniles/100 ml of soil in the initial inoculation treatment showed that it was able to reduce kenaf production (Supriyono & Suhara, 2007) and reduce kenaf fiber quality (Dalmadiyo et al., 1989). Plant damage due to root-knot nematodes increases with the number of nematodes infecting root tissue.

The use of resistant varieties is an appropriate way to suppress nematode population density and limit the damage threshold, thereby reducing crop yield losses (de Deus Barbosa et al., 2009; Davis & Stetina, 2016; Costa et al., 2017). Before assembling kenaf varieties that have resistance to NPA, it is very important to know about the information on genes controlling resistance to NPA.

^{*} Corresponding Author: nikicro@yahoo.co.id

Inheritance of kenaf resistance to root-knot nematodes until now there is not enough information. Several studies have been carried out on several plants to determine the genes controlling resistance to NPA. Some of the results of research on plant resistance to NPA include; Rosella resistance to NPA is controlled by monogenic genes with dominant gene action (Falusi, 2008); cotton resistance to NPA is controlled by two dominant genes (McPherson et al., 2004) and resistance of tobacco plants to NPA M. incognita race 2 is additive (Shahadati et al., 2017). The values of heritability in the broad sense and heritability in the narrow sense for the number of root cavities were 0.93 and 0.75, respectively, the number of egg masses 0.94 and 0.56 and the number of eggs per egg mass were 0.61 and 0.28, respectively. The research results of Wang et al. (2017) that the resistance of cotton to root-knot nematodes is controlled by chromosome 11. QTL 11 has a negative effect on the number and production of nematode eggs.

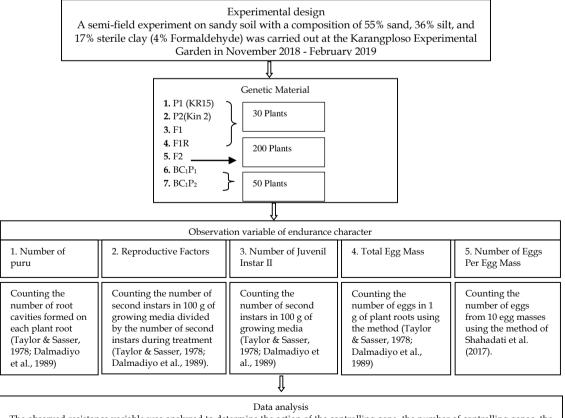
Analysis of the inheritance of kenaf resistance to NPA *M. incognita* has been carried out by several researchers by utilizing the source of resistance from close relatives of kenaf that have resistance to NPA *M. incognita,* including *H. sabdariffa, H. acetosella* and also *H. radiatus.* The availability of this information has not been

able to provide much progress in breeding the resistance of NPA *M. incognita*. This is because there are constraints in the propagation of material for further study. Several obstacles in the inheritance of kenaf resistance to NPA *M. incognita* were caused by the high *incompatibilities* and sterility of the crosses between kenaf and its close relatives.

There is not enough information on the inheritance of resistance traits in intraspecific kenaf crosses. This is because the source of the gene for resistance to NPA in kenaf has not been obtained. Based on several research results, it is stated that the highest resistance of kenaf to NPA is tolerant (Yulianti & Supriyono, 2009). This has an impact on breeding methods which generally require a relatively long time because they must collect tolerant genes to obtain resistant genotypes.

Based on this, it is necessary to conduct a study on the inheritance of kenaf resistance to NPA in order to determine an effective and efficient breeding program strategy to obtain high yielding and NPA resistant kenaf genotypes. The purpose of this study was to estimate the inheritance of kenaf (*H. cannabinus L*) resistance to NPA *M. incogita*.

Method



The observed resistance variable was analyzed to determine the action of the controlling gene, the number of controlling genes, the estimated value of broad heritability, narrow heritability and the presence or absence of maternal effects.

Maternal effects were tested by comparing the mean values of F1 and F1R with the t-test according to (Daryanto, 2016; Hapshoh, 2016) at the 5% level. The frequency distribution of the F2 population was tested by normality test using the Kolmogorov-Smirnov method.

The degree of dominance is calculated based on the formula of Petr & Frey (1966) namely hp = (F1 – MP) / (HP – MP); with *hp* = potential ratio, F1 = average value of F1, HP = average value of the highest elder, MP = middle value of the two elders. Based on the ratio potential value, the degree of dominance is classified as: HP = 0: no dominance, HP = 1 or HP = -1: fully dominant or recessive, 0 <HP< 1: partial dominant (incomplete positive dominant), -1 < hp < 0: partially recessive (imperfectly negative dominant), and HP > 1 or hp < -1: over dominant.

Estimation of the number of controlling gene pairs for each nematode resistance character used the calculation described by Lande (1981) namely N=µ_{P1}µ_{P2}/8(σ_{F2}^2 -2(σ_{F1}^2 + σ_{P1}^2 + σ_{P2}^2)/4 where µ_{P1} = Average of elders, Kin2, µ_{P2} = Average range of elders KR15, σ_{F2}^2 = Variety of F₂, σ_{P1}^2 = Variety of elders Kin2 σ_{P2}^2 = Variety of elders KR15. Heritability in broad sense, calculated based on the formula (Acquaah, 2007) i. e. h²_{bs} = σ_{F2}^2 -1/3(σ_{P1}^2 + σ_{P1}^2)/ σ_{F2}^2 where h²_{bs} = broad sense heritability, σ_{F2}^2 = F2 variety, σ_{P1}^2 = Kin2 parent variety, σ_{P2}^2 = KR15 parent variety and σ_{F1}^2 = F1 variety.

Narrow sense heritability is calculated based on the formula (Syukur et al., 2015) which is $h_{ns}^2 = 2\sigma_{F2}^2 - (\sigma_{BC1P1}^2 + \sigma_{BC1P2}^2) / \sigma_{F2}^2$. Where $h_{ns}^2 =$ narrow sense heritability, $\sigma_{BC1P1}^2 =$ population variance BC1P1, $\sigma_{BC1P2}^2 =$ population variance BC1P2, $\sigma_{F2}^2 =$ population variance

F2. The estimated heritability value is considered low if $h^2 < 20\%$, moderate if $20\% \le h^2 \le 50\%$ and high if $h^2 > 50\%$

The feasibility of the additive and dominant models was estimated by three genetic parameters, namely the mean (m), the amount of influence of the additive gene (d) and the amount of influence of the dominant gene (h) using the Joint Scaling Test method (Mather & Jink, 1982). If the action of the gene does not meet the additive-dominant model, a test is carried out to determine whether there is a non-allelic gene interaction using an epistatic model with six parameters (Mather & Jink, 1982).

Result and Discussion

The influence of the female elder

Tests for the influence of female parents were carried out to determine whether the resistance of kenaf to NPA *M. incognita* was influenced by female parents. To determine whether there was an influence of female parents on the resistance of kenaf to NPA *M. incognita*, a t-test was carried out on the F1 population with reciprocity. Based on the results of the comparison between the F1 population average and its reciprocity, it shows that there is no reciprocal effect as indicated by the probability value of t which is greater than 0.05.

The results of the homogeneity test of the resistance to root-knot nematodes using the F test on Kin2 x KR15 showed that the F1 and F1R variants were homogeneous. This can be seen from the calculated F value which is greater than the F table at the 5% level (Table 5). Thus, in the next analysis, the F1 and F1R data can be combined as F1 population data.

Table 1. The average value and standard deviation of the kenaf resistance character against the nematode M. incognita in F1 and F1R, the results of the test of different mean values and homogeneity of variance.

incognita in i	incognita in FF and Firly the results of the test of anterent mean values and nontogenenty of variance.								
Population	IP	FR	J2	JMT	JTMT				
F ₁	70.27±3.48	5.41±0.01	35.77±0.65	11.73±0.45	402.43±9.45				
F _{1R}	72.7±6.2	5.71±0.36	34.01±1.60	12.87±1.30	403.8±16.0				
Prob>(t)	0.19tn	0.55tn	0.62tn	0.33tn	0.84tn				
Prob>(F)	0.93tn	0.70tn	0.73tn	0.73tn	0.77tn				

Information: JP = number of shells, FR = reproductive factor, J2 = number of 2nd instar juveniles, JMT = number of egg masses and JTMT = number of eggs per egg mass. tn = not significantly different

The character of kenaf plant resistance to NPA *M. incognita* was not affected by female parents (Table 1). This indicates that the resistance trait is controlled by *the nuclear genes.* (Hulupi, et al., 2007) stated that the resistance of Arabica coffee to *Radopholussimilis Cobb* showed that it was not influenced by female parents. The resistance properties of kenaf to NPA *M. incognita* showed similar to the resistance properties of chili plants to anthracnose or to aphids. According to Syukur et al. (2007); Hapshoh (2016) that the resistance of chili plants to anthracnose disease is not influenced by female parents. Daryanto (2016) stated that the resistance of chili to aphids was also not influenced by female elders.

Analysis of the inheritance of resistance characters

Analysis of the inheritance of kenaf resistance characters against the nematode *M. incognita* included: normality test, degree of dominance and number of controlling genes, variance components, genetic model feasibility and heritability values.

Normality Test of Population Frequency F2

Analysis of the normality of the data in F2 was used to determine the number of genes controlling the resistance character of *M. incognita NPA*. The results of the normality test on all variables of kenaf plant resistance to NPA *M. incognita* showed that the F2 population frequency distribution was not normal (P < 0.10) (Table 2).

Wanda et al. (2014) stated that the distribution of F2 population data was not normal indicating that the character was controlled by a simple gene or a major gene. Furthermore (Barnawi et al., 2013) explained that the qualitative character of a plant is usually controlled by one or two genes, less influenced by the environment, so it is relatively easy to handle in breeding. The segregation pattern of the qualitative characters follows the Mendelian ratio and its modifications.

Table	2. The resu	alts of the	normali	ty ana	lysis of	the
kenaf	resistance	variable	against	NPA	using	the
Kolmo	gorov-Smir	nov test.				

Resistance	Statistic	Df	Augraga	Value of
Variable	Statistic	DI	Average value	р
JP	3.22	200	71.77±6.62*	0.000†
FR	2.98	200	4.94±2.89*	0.000†
J2	1.56	200	38.99±1.37*	0.015†
JMT	2.87	200	9.50±4.96*	0.000†
JTMT	3.71	200	4.68±1.65*	0.000†

Note: Number of Puru (JP); Reproductive Factors (FR); Number of Juvenil Instar 2 (J2); Total Egg Mass (JMT); Number of Eggs per Egg Mass (JTMT) *= mean \pm SD; = p<0.05 data is not normally distributed

Degree of Dominance and Number of Controlling Genes

Estimation of the degree of dominance was carried out by estimating the potential ratio (*hp*) of the median value of the two parents and F1 using the calculations described by Petr & Frey (1966). The potential value of the ratio of the results of the Kin2 x KR15 cross from the number of shells, reproductive factors, juvenile instar 2, number of egg mass and number of eggs per egg mass were 0.05, 0.05, 0.07, 0.15 and 0.01, respectively (Table 3).

Table 3. Value of	potential ra	atio and nun	nber of genes	s controlling l	kenaf resis	tance to NPA	M. incognita.

	The mean value of the resistance variable									
Population	JP	FR	J2	JMT	JTMT					
P ₁	109.47±4.96	8.89±0.01	55.90±0.65	19.61±0.65	561.98±8.65					
P ₂	34.80±3.32	2.26±0.01	22.16±0.42	5.89±0.32	282.21±7.32					
F ₁	70.27±3.48	5.41±0.01	37.77±0.65	11.73±0.45	390.58±9.45					
MP	72.08	5.57	39.03	12.75	391.51					
hp	0.05	0.05	0.07	0.15	0.01					
Gene action	Partially positive c	lominant								
Number of genes	0.01	0.12	0.03	0.03	0.02					

Note: JP: number of purua kar, FR: reproductive factor, J2= number of juveniles in the 2nd instar, JMT: number of egg masses, and JTMT: number of eggs per egg mass.

The degree of dominance (hp) on all of the evaluated resistance variables ranged from 0.03 to 0.55 (Table 3). If the hp value of a character is in the range of 0 and 1, it indicates that the character is controlled by the action of an imperfectly positive dominant gene (partially positive dominant). This agrees with the results of research by Setyo-budi (2009) which states that the inheritance of the kenaf genotype resistance to rootknot nematodes is controlled by the role of partially positive dominant genes. Bertrand et al. (1995) stated that the action of genes controlling resistance to the nematode Meloidogyne spp. that infects Catimor Arabica coffee is the action of a partially dominant gene. Hulupi et al. (2007) added that the resistance of Arabica coffee to Radopholussimilis Cobb was controlled by a partially dominant gene. According to Roberts et al. (1998) resistance of tomato varieties to Meloidogyne spp on the character of the number of egg masses and the rate of formation of the bladder was controlled by genes with partially dominant gene action. The degree of partially positive dominance in the F1 genotype from the Kin2 x KR15 cross indicated a strong gene dominance event, and the inheritance was controlled by partial dominance.

Estimation of the genetic component

To determine the action of genes controlling the resistance variable to M. incognita NPA, a combined scale test was performed (Mather & Jink, 1982). Individual scale testing compares the t-count value with the t-table to state whether or not a genetic component is real (Singh & Chaudhary, 1979). Estimation values of genetic parameters of individual scale test (scaling test) of kenaf resistance variable to NPA M. incognita are presented in (Table 3).

			parameters					

Parameter	JP	FR	J2	JMT	JTMT
m	398.4	28.25	207.11	57.63	2233.11
[d]	37.33	3.32	16.87	6.86	109.30
[h]	6.99	1.34	2.84	2.84	101.48
C	265.21	21.01	129.39	39.85	1305.03
SE. C	193.43	13.83	67.09	43.70	671.35
t-count	1.37tn	1.52tn	1.93tn	0.82tn	1.94tn
t-table	1.96	1.96	1.96	1.96	1.96

C = 4 (mean F2) - (mean P1) - 2 (mean F1) - (mean P2) (Singh and Chaudary, 1979); m: mean value, d: additive value, h: dominance value, C: gene interaction value, SE C: standard error C. IP=Number of root cavities, FR= Reproductive factors, I2= number of 2nd instar juveniles, JMT=Number of egg masses and JTMT= Number of eggs per egg mass. ** = significant at 1% t-test level = 2.58 * = significant at 5% t-test level = 1.96, tn = not significant

Based on (Table 4) shows that the appropriate genetic model for the variable resistance to NPA M. incognita on all variables evaluated in the Kin2 x KR15 cross is the additive-dominant model (m[d][h]). This model is the most suitable model, this is because the value of t-count is smaller than t-table = 1.96.

The character of kenaf plant resistance to NPA M. incognita was controlled by one gene. This agrees with the results of Falusi (2008) which states that the resistance of rosella to SSP nematodes is controlled by one gene. Meanwhile, Shahadati et al. (2017) who conducted research on tobacco resistance to M. incognita NPA stated that tobacco plant resistance to M. incognita NPA was controlled by one gene, with a partially dominant effect. The number of controlling genes indicates the number of genes that are effective in controlling the expression of a character. The most suitable gene action model for the character of kenaf resistance to NPA M. incognita was additive-dominant (m[d][h]). Thus, it means that only additive [d] and dominant [h] gene action determines the diversity of kenaf resistance to NPA M. incognita. The value of the

additive variance is greater than the dominant variance, this indicates that the genetic variance is determined by the action of the additive gene.

The visible variety of appearance of a plant (phenotype) is the result of a combination of diversity due to genetic and environmental influences. The genetic diversity that is the focus of plant breeding is the diversity caused by the influence of additive gene action, the influence of dominant gene action and the influence of interactions between genes (Falconer, 1960).

Heritabilitas

38.05

66.69

61.21

91.78

Heritability is a reflection of the influence of genes (genotype) on the observed external appearance (phenotype). Heritability values in the broad sense (h2bs) and narrow sense (h2ns) on the results of the Kin2 x KR15 cross for the kenaf resistance variable to NPA M. incognita which included the number of lungs, reproductive factors, number of juveniles instar 2, number of egg masses and average number eggs per egg mass was high (Table 5).

Table 5. Variance components a	and heritability values of	kenat resistance	e to the nematod	le M. incognita.	
Genetic parameters	JP	FR	J2	JMT	JTMT
Environmental Variety	881.32	3.54	50.95	38.70	3503.12
Variety of Additives	7628.57	41.91	1016.20	401.28	107262.94
Dominant Variety	4760.56	20.02	506.31	215.03	23303.85
Variety of Phenotypes	5885.74	29.50	685.63	293.10	62960.55

Table E Vari

7957.20

56.11

51.71

92.15

Description: JP = number of shells, FR = reproductive factors, J2 = number of juveniles instar 2, JMT = number of egg masses and JTMT = average number of eggs per egg mass.

High additive effect has an impact on the effectiveness of the selection. In plant breeding, the selection of traits controlled by additive genes is expected to get a large and rapid selection progress. Selection of characters with high heritability can be done in the early generations. (Barmawi et al., 2013) said that a character that has a high heritability value can be

Variety of BC₁P₁ and BC₁P₂

Proportion h_{ns}^2/h_{bs}^2 (%)

 $h_{bs}(\%)$

 $h^2ns(\%)$

selected in the early generations (F2 and F3). The high value of heritability in the broad sense and in the narrow sense for the resistance of NPA M. incognita would be a good step because in the selection program high selection progress would be obtained.

385.55

63.01

57.86

91.83

863.16

80.44

56.31

70.00

The total phenotypic appearance seen from the heritability value in the broad sense of the variable number of shells (56.11%), reproductive factors (66.69%),

72289.63

86.95

81.56

93.80

number of juvenile instar 2 (80.44%), total egg mass (63.01%) and average number of eggs per egg mass (86.95%) was influenced by genetic variation. In all variables of resistance evaluated, it was 51.71% on the number of shells, 61.21% on the reproductive factor variable, 56.31% on the variable number of juvenile instars 2, 57.86% on the variable number of egg masses and 81.56% on the variable average number of eggs per egg mass of the total variance was influenced by additive genes, and there was a difference of 4.40% in the number of shells, 5.48% in reproductive factors, 24.13% in the number of juvenile instars of 2, 5.15% in the number of egg masses and 5.39% in the average number of eggs per egg mass. This indicates the small role of the influence of non-additive gene action (dominant and interactions between genes) on the character of kenaf resistance to NPA M. incognita.

The magnitude of the influence of genetic variance from the total phenotypic appearance on all evaluated resistance variables indicated the small role of the influence of environmental variation (E=43.89% for the number of shells, 33.31% for reproductive factors, 19.56 for the number of juvenile instars 2, 36.99% for egg mass and 13.05% for the number of eggs per egg mass). This is consistent with the results of the estimation of the components of genetic variance that the environmental variance components have no effect on all resilience variables (Table 5).

The basis for the success of plant breeding work is largely determined by the amount of genetic diversity that can be passed from parents to offspring. The additive component is the only component of genetic variation that is passed from parents to offspring, and the narrow heritability value again shows this magnitude through the genetic proportion of total appearance as the portion that is completely passed on from parents to offspring (Falconer 1960; Poehlman & Sleper, 1995). From the point of view of breeding kenaf, which is a self-pollinating plant, selection should be made for additive effects in the hope of gathering superior genotypes; selection becomes ineffective if the superior genotype is determined by the influence of dominance and interactions between genes (Poehlman and Sleper, 1995).

Conclusion

Based on the results of the study, it can be concluded that the resistance of kenaf to NPA *M. incognita* is controlled by the genes present in the cell nucleus. Kenaf resistance to NPA *M. incognita* was controlled by a dimonan gene with heritability values ranging from 56.11 to 86.95.

Acknowledgements

We would like to thank the Agency for Agricultural Research and Development – Ministry of Agriculture of the Republic of Indonesia which has provided funding for this research with research contract number 01/SPK/120.001/VII/2019 dated July 31, 2019. We also thank my friends from Faculty of Agriculture – Universitas Brawijaya (Farhan Amin Gemayel and Naufal Zaki).

References

- Acquaah, G. (2007) Principles of Plant Genetics and Breeding. 1st edn, Blackwell. 1st edn. Edited by G. Acquaah. USA: Blackwell Publishing.
- Adegbite, A.A. (2018). Response of selected kenaf cultivars to Meloidogyne incognita under greenhouse conditions. International Journal of advance Agricultural Research, 6(-), pp. 55–58. <u>http://dx.doi.org/doi.org/10.33500/ijaar.2018.</u> 06.006
- Barmawi, M., Yushardi, A. & Sa'diyah, N. (2013). Daya waris dan harapan kemajuan seleksi karakter agronomi kedelai generasi F2 hasil persilangan antara yellow bean dan Taichung. *Journal Agrotek Tropika*, 1(1), pp. 20–24. http://dx.doi.org/10.23960/jat.v1i1.1882
- Bertrand, B.; F. Anzueto; M.X. Pena; F. An- thony & A.B. Eskes (1995). Genetic improvement of coffee for resistance to root-knot nematodes (Meloydogyne spp.) in Central America. ASIC, 16 e Colloque, Kyoto, 630 – 636.
- Costa, G.A.S.H., Silva, V.B.G., Marc Augusto, Vianna, P.B.R. & Fabrício. (2017). Inheritance of resistance to *Meloidogyne incognita* race 3 in cotton accession TX 25. *Acta Scientiarum*, 39(3): 331–337. <u>http://dx.doi.org/10.4025/actasciagron.v39i3.325</u> <u>63</u>.
- Dalmadiyo, G., Pusposendjojo, N. & Rahayu, B, T. (1989). Effect of initial population densities of root knot nematode (Meloidogyne spp) on growth and yield of kenaf (H. cannabinus L.). *BPPS UGM*, 2(1B), pp. 165–176.
- Daryanto, A. (2016). Analisis Genetik dan Pewarisan Sifat Ketahanan Cabai Terhadap Infestasi Kutu Daun MelonAphis gossypii Glover (Hemiptera: Aphididae). Tesis. Fakultas Pertanian. Institut Pertanian Bogor.
- Davis, R.F. & Stetina, S. R. (2016). Resistance and tolerance to nematodes in cotton', in Galbieri eJean, R. L. B. (ed.) Nematoides fitoparasitas do algodoeiro nos cerrados brasileiros: Biologia e medidas de controle. 1st edn. Cuiaba: Dados Internacionais de Catalogação na Publicação (CIP) Instituto, pp. 1–242.

- deDeus Barbosa, A.E.A., Rocha Fragoso, R, deLimae Souza, D.S., Freire, E, de Oliveira Neto, O.B., Barbosa, A.A.V., Coiti, R.T., Guimara~es, L.M., Martins, N.F., Cia, E, Fernandez, D, de Lima, L.M., Silva, M.C.M., Rocha, T.L. & Grossi-de-Sa, M.F. (2009). Differentially expressed genes in cotton plant genotypes infected with Meloidogyne incognita. Plant Science, 177: 492-497. http://dx.doi.org/10.1016/j.plantsci.2009.07.013.
- Falconer, D.S. (1960). *Introduction to Quantitative Genetics*.1st edn. Edited by D. Falcorner. Londo: Printed in Great Britain for Oliver and Boyd Ltd.
- Falusi, O. A. (2008). Inheritance of characters in kenaf (Hibiscus cannabinus). *African Journal of Biotechnology*, 7(7), pp. 904–906.
- Falusi, O.A. (2008). Inheritance of characters in kenaf (Hibiscus cannabinus). *African Journal of Biotechnology*, 7(7), pp. 904–906. <u>http://dx.doi.org/10.5897/AJB07.694</u>.
- Hapshoh, S. (2016). Pewarisan Karakter Kualiatatif dan Kuantitatif pada Persilangan Cabai Besar dan Cabai Rawit serta Ketahannya terhadap Penyakit Layu Fusarium. Tesis. Fakultas Pertanian. Institut Pertanian Bogor.
- Hulupi, R., Nasrullah & Soemartono (2007). Inheritance of Arabica Coffee resistance to Radopholus similis Cobb. *Pelita Perkebunan*, 23(1), pp. 1–16. <u>https://doi.org/10.22302/iccri.jur.pelitaperkebun</u> <u>an.v23i1.53</u>
- Lawrence, G. & McLean, K. (1992). Host status response of kenaf (H.cannabinus) to Meloidogyne incognita race 4, M. javanica, Hoplolaimus magnitylus, and Rotylenchulus reniformis. *Nematropica*, 22(2), pp. 247–250. Retrieved from <u>https://agris.fao.org/agris-</u> search/search.do?recordID=US9323446
- Mather, S.K. & Jink, J.L. (1982). *Biometrical Genetic*. 3rd edn. Edited by S. K. Mather and J. L. Jink. Birmingham: Springer Science Business Media. Retrieved from https://link.springer.com/book/10.1007/978-1-4899-3404-8
- McPherson, M.G., Jenkins, J.N., Watson, C.E. & McCarty, J.C. (2004). Inheritance of Root-knot nematode resistance in M-315 RNR and M78-RNR cotton. *Journal of Cotton Science*, 8(3): 154–161.
- Petr, F. C., & K.J.Frey. (1966). Genotipe correlations, dominance and heritability of quantitative charachters in oats. *Crop Science*, 6(3), 259–262. https://doi.org/10.31274/rtd-180813-2994
- Poehlman, J.M. & Sleper, D.A. (1995). *Breeding Fields Crops.* 4th edn. Edited by J. M. Poehlman and A. S. David. Iowa: Oxford and IBH Publishing.
- Roberts, P.A, W.C. Matthews & J.C. Veremis (1998). Genetic mechamisms of host- plant resistance to

nematodes. p. 209 – 238. In : Barker K.R., G.A. Pederson & G.L. Windham (eds.) Plant and Nematode Interactions. Agronomy Monograph no. 36. Madison, Wiscon- sin, USA

- Setyo-Budi, U., Sudjindro & Dyah, R. (2009). Resistance variability of kenaf (Hibiscus cannabinus L.) genotypes to root knot nematode (Meloidogyne incognita). *Jurnal Litri*, 15(2), pp. 60–65.
- Shahadati-moghaddam, Z., Bagheri, N. & Jelodar, N.B. (2017). Inheritance of southern root-knot nematode resistance in air-cured tobacco. *Tropical Plant Pathology*. Tropical Plant Pathology, 42, pp. 32–38. <u>http://dx.doi.org/10.1007/s40858-016-0122-4</u>.
- Supriyono & Suhara, C. (2007). Evaluasi ketahanan plasma nutfah kenaf dan kerabatnya terhadap Fusarium sp. dan nematoda puru akar Meloidogyne spp. Malang.
- Syukur, M., Sriani and Yunianti, R. (2015). *Teknik Pemuliaan Tanaman*. 2nd edn. Edited by S. Syukur. Bogor: Penebar Swadaya. Retrieved from: <u>https://books.google.co.id/books?id=icNUCgAA</u> <u>QBAJ&pg=...false</u>.
- Syukur, M., Sujiprihati, S. & Koswara, J. (2007). Inheritance of Resistance to Anthracnose caused by Colletotrichum acutatum in Pepper (Capsicum annuum L.). *Bul. Agron*, 35(2), pp. 112–117.
- Tahery, Y, Aini, N.A.S., Abdul-Hamid, H, Puad, M.A. & Norlia, B. (2011). Status of root knot nematode disease on kenaf cultivated on bris soil in kuala terengganu. *World Applied Sciences Journal*, 5(9): 1287-1295.
- Taylor, A. & Sasser, J. (1978). Biology, Identification and Control of Root-knot Nematodes (Meloidogyne species) Raleigh. 1st edn, NC: United States Agency for International 1st edn. Edited by A. Taylor and J. Sasser. North Carolina: North Caroliba State University Graphics. Retrieved from http://scholar.google.com/scholar?hl=en&btnG= Search&q...(Meloidogyne+Species)#5.
- Wanda, N., Barmawi, M. & Akin, H. M. (2014). Pola segregasi karakter ketahanan tanaman Kedelai (Glycine max [L]. Merrill) terhadap infeksi soybean mosaic virus populasi F 2 keturunan Taichung X Tanggamus. *Jurnal Penelitian Pertanian Terapan*, 15(1), pp. 54–60.

https://doi.org/10.25181/jppt.v15i1.112

- Wang, C, Ulloa, M, Duong, T.T. & Roberts, P.A. (2017). QTL analysis of transgressive nematode resistance in tetraploid cotton reveals complex interactions in chromosome 11 regions. *Frontiers in Plant Science*, 8:1-12. <u>https://doi.org/10.3389/fpls.2017.01979</u>.
- Wokoma, E., Tanimola, A. & Chizzy, M. (2015).
 Evaluation Kenaf (H cannabinus) Accession for Resistance to Meloidogyne incognita. *International Journal of Development Research*, 5(3), pp. 3645–3650.
 Retrieved from 697

https://www.journalijdr.com/evaluation-...incognita

Yulianti, T. & Supriyono (2009). Penyakit tanaman kenaf dan pengendaliannya. in Sulistyowati, E. (ed.) *Monograf Balittas Kenaf*. 1st edn. Malang: IAARD Press, pp. 93–105.