



Antidiabetic Molecular Mechanisms of Active Compounds from Several Orchids

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Abstract: Hyperglycemia condition that leads to diabetes causes various complications. Various active compounds from plants have been studied for their antidiabetic abilities. One of them is the orchid plant. Besides being used as decoration, orchids contain several active compounds that have been proven to be used in medicine, including diabetes. This article discusses the antidiabetic mechanism of several active compounds obtained from orchids. Publication regarding orchid plant for diabetes were found in databases such as PubMed, Google Scholar, Wiley, Science Direct, Medline, Scopus, and Springer. Keywords used in this study were “orchid”, “diabetes”, “hyperglycemia”, “compound” and “herbal”. Out of the 447 collected articles (published in the period between 2011 and 2022), 416 were excluded due to non-relevant studies. There were 31 eligible studies included in this article. In conclusion, the antidiabetic mechanisms of the orchid extracts were as antioxidant, anti-inflammatory and anti-glycation agents, increasing insulin action, influencing lipid metabolism, and inhibiting α -amylase and α -glucosidase activity.

Keywords: Active compound; Antidiabetic; Mechanism; Orchid

Introduction

Hyperglycemia in diabetics can increase the generation of Advanced Glycation End Products (AGEs), a glycated protein molecule whose interaction with its receptor, RAGE, can induce the formation of Reactive Oxygen Species (ROS) through NADPH oxidase activation. One example of AGEs is H_2O_2 , of which its excessive production can activate nuclear factor kappa B (NF κ B) and further causes diabetes complications (Yulianti et al., 2021, 2022). There are some studies of medicinal plants that were traditionally used to treat diabetes. Their antidiabetic activities such as improved insulin sensitivity and hypoglycemic activities. These abilities are believed due to their phenolic, flavonoids, terpenoids, alkaloids, and glycosides compounds (Salleh et al., 2021). Some plants have antioxidant abilities and are used as effective herbal medicines. While their antidiabetic compounds

increase insulin secretion of pancreatic tissues or decrease the intestinal absorption of glucose (Kooti et al., 2016).

Orchids are member of Angiosperm group and belong to Orchidaceae family (Tsering et al., 2017). There are 4000 species of orchids found in Indonesia (Martha & Rahayu, 2022). Orchidaceae is a flowering plant comprises of 25,000 to 35,000 species from 750 to 850 genera (Musharof Hossain, 2011). Orchids are usually used preferably for decorative flower rather than for its medicinal use. Chemical compounds from this species include polysaccharides, reveral, stilbenoids, while the polyphenol compounds are alkaloids, flavonoids, terpenoids, bibenzyl derivatives and phenanthrenes (Ashu Rajeshbhai & Ingalhalli, 2022; Joshi et al., 2020; Martha Pérez Gutiérrez, 2010; Singh et al., 2012). Orchids has the ability of antidiabetics, anti-inflammation, antioxidant, antimicrobes, anticancer, neuroprotection, and antiallergy (Ashu Rajeshbhai &

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Ingalhali, 2022; Minh et al., 2016). Minh et al. (2016) found that some compounds, from hybrid *Phalaenopsis* spp. namely phenolic, vanillic acid, protocatechuic acid, caffeic acid, p-hydroxybenzoic acid, syringic acid, ferulic acid, vanillin, p-coumaric acid, sinapic acid, benzoic acid, and ellagic acid have potentials as antioxidants. Orchidaceae, as a big plant family, is well known as a source of bioactive compounds, which has been studied for its new bioactive natural products (Kuo et al., 2022). This article reviews some orchids with their medicinal properties, which can be used as antidiabetics along with their particular mechanisms, using the most recent investigations available on literatures. Many studies have been carried out on the benefits of orchids for the treatment of diabetes, but have not discussed all the possible mechanisms that can be carried out by the active compounds contained in these orchids.

Method

Scientific databases such as Science Direct, PubMed, Google Scholar, Medline, Wiley, Scopus, and Springer were utilized to find publication regarding orchid plant for diabetes. The keywords used in this study included "orchid", "diabetes", "hyperglycemia", "compound" and "herbal". Out of the 447 collected articles (published in the period between 2011 and 2022), 416 were excluded due to non-relevance studies.

Inclusion and Exclusion Criteria

The search was restricted to only articles written in English language. All studies found during the search were evaluated by three different authors independently. After compliance with inclusion criteria, the eligibility screening was done by reading title and abstract of all articles to remove irrelevant studies. Most of the unselected journals were due to the plants studied do not belong to the Orchidaceae, or even though they are known as orchid trees or orchid, but they have no antidiabetic activities. Some of the journals were the same journal, or not in English. There were ultimately 31 eligible studies that included in this article. The data extraction was performed by classifying the articles to answer the research questions. Research design and method should be clearly defined.

Result and Discussion

Inclusion Studies on Antidiabetics Mechanism of Orchids Extract

There are several types of orchids that have been found to demonstrate antidiabetic abilities. The antidiabetic ability shown by these orchids are through various mechanisms (Table 1 Antidiabetes properties of

some orchids and heir mechanisms). The antidiabetic activity of orchids were found using different extraction methods, plant parts and research methods (*in vitro* and *in vivo*).

Table 1 shows various antidiabetic activities possessed by the compounds found from the extraction of various plant parts from orchids. Almost all parts of the plant can be used and have antidiabetic activity. The parts of the orchid plants used in these studies included flower parts, leaves, stems, roots, and even some studies used all parts of the plant. The active compounds were obtained using various solvents, namely chloroform, ethanol, methanol, hexane, ethyl acetate and water. This shows that the active compounds produced in these studies have different polarities, from polar to non-polar. Solvent polarity affected phytochemical substances obtained from the isolation and purification steps of plant material. Phytochemicals are extracted in solvents of different polarity as no single solvent (Nawaz et al., 2020). There are several antidiabetic mechanisms carried out by active compounds extracted from orchids, which are as follows:

Antioxidant Activity

The extract of some orchids, *Dendrobium huoshanense*, *D. officinale*, *D. nobile*, *D. chrysotoxum*, *P. karwinskii*, *Prosthechea michuacana* (Lex.) W.E. Higgins, *Dendrobium crepidatum*, *Dendrobium aqueum* Lindl, *Dactylorhiza romana* subsp. Georgica, *Dendrobium officinale*, *Anacamptis pyramidalis* (L.) Rich, *Bauhinia variegata*, *Gastrodia elata* Blume, *Maxillaria tenuifolia*, *Eulophia ochreatea* L, *Gymnadenia orchidis* Lindl and *Grammatophyllum speciosum* Blume have an activity as antioxidant. The antioxidant activities were done by controlling and scavenging free radicals to offer cell protection against oxidative stress (Gutierrez, 2013; Hunyadi, 2019) as well as acting as antioxidant via their hydrogen-donating ability (Paudel et al., 2019). They are capable of donating hydrogen to a free radical in order to remove odd electron, which is responsible for the radical's reactivity (Mukherjee et al., 2012; Nimse & Pal, 2015). Many of compounds have antioxidant properties which act through either enzymatic or non-enzymatic pathways (Safari et al., 2018). Table 1 shows several species of orchids containing flavonoids. Flavonoids as antidiabetic compounds act as antioxidants because they have three benzene rings and one hydroxyl group (Ahmed et al., 2020; Yeh et al., 2017). The antioxidant activity of phenolic compounds varies, their biological activity depends on the structure of these compounds, their combination with other compounds, their solubility, absorption and metabolism (Francenia Santos-Sánchez et al., 2019; Ji et al., 2020).

Table 1. Antidiabetes Properties of Some Orchids and Heir Mechanisms

Orchid	Part of the plant	Model description	Major finding	Control	Isolated Bioactive compound	Reference
<i>Dendrobium huoshanense</i> (DHP), <i>D. officinale</i> (DOP), <i>D. nobile</i> (DNP) and <i>D. chrysotoxum</i> (DCP)	freeze-dried stems	alloxan induced diabetic Male Kunming mice	hypoglycemic and antioxidative activities between four <i>Dendrobium</i> polysaccharides	metformin	polysaccharides	(Pan et al., 2014)
<i>Prosthechea karwinskii</i>	leaves extract (100, 200, and 300 mg/kg p.o.)	Metabolic syndrome was induced in Wistar rats through administration of a 40% sucrose diet for 20 weeks	decreasing weight gain, abdominal and pericardial fat deposits, insulin resistance, triglycerides, TNF- α , HS-CRP, and adiponectin	metformin (200 mg/kg p.o.)	quinic acid, neochlorogenic acid, chlorogenic acid, rutin, kaempferol-3-o- β -rutoside, and embelin	(Barragán-Zarate et al., 2021)
<i>Prosthechea karwinskii</i> (Mart.) J.M.H. Shaw	leaves	inhibition of ROS using peripheral blood mononuclear cells (PBMC). Inflammation using Female Wistar rats induced 100 μ L of carrageenan 1% (w/v)	inhibiting reactive oxygen species and exerted an anti-inflammatory effect	naproxen	quinic acid, malic acid, neochlorogenic acid, chlorogenic acid, rutin, embelin, pinellic acid, and azelaic acid	(Barragán-Zarate et al., 2020)
<i>Prosthechea michuacana</i> (Lex.) W.E. Higgins (PM)	bulbs hexane extract	streptozotocin (STZ) and nicotinamide-induced type 2 diabetic male CD1 mice	anti-diabetic effect by stimulating insulin -dependent and by protecting pancreatic β -cells from oxidative stress and also an anti-obese, anti-insulin resistance and antihyperglycemic pro-drug.	glibenclamide	triterpenes (tetracyclic triterpenoids: 24-methyl, 24-hydroxy-5 α -lanosta-9(11), 25-dien-3 α -acetate and 24-methyl-24-hydroxy-5-lanosta 9 (11)-en-3 α acetate)	(Gutierrez, 2013)
<i>Dendrobium crepidatum</i>	Ethanol and acetone stem's extracts	DPPH (2,2-diphenyl-1-picrylhydrazyl), in vitro	inhibiting DPPH free radicals	ascorbic acid	tetracosane, triacontane, stigmasterol, and some phenol derivatives (2-methoxy-4-vinylphenol, 2-methoxy-5-(1-propenyl)-phenol, p-mesyloxyphenol, and 2,6-dimethoxy-4-(2-propenyl)-phenol)	(Paudel et al., 2019)
<i>Malaxis rheedei</i> SW	whole plant methanol extract	α -amylase and α -glucosidase activities with	inhibiting the enzymes like salivary, amylase and glucosidase and shows potential		flavonoid, tannin, glycoside, resin, steroids,	(Haridas et al., 2017)

Orchid	Part of the plant	Model description	Major finding	Control	Isolated Bioactive compound	Reference
		half-maximal inhibitory concentration (IC50), in vitro	activities against diabetes mellitus disease		terpenoids, cardiac glycosides and triterpenoids etc	
<i>Dendrobium aqueum</i> Lindl	aqueous extracts of pseudobulbs	Antioxidant activity using DPPH, the protein glycation inhibitory using albumin/fructose glycation model	DPPH free-radical scavenging potential and antiglycation potential			(Mukherjee et al., 2012)
<i>Dactylorhiza romana</i> subsp. Georgica	tubers chloroform, ethyl acetate, methanol, ethanol and hexane extract	phosphomolybdate test method, DPPH scavenging activity, α -amylase and α -glucosidase activity	antioxidant and antimicrobial agent, or enzyme inhibitor		Gallic acid, protocatechuic acid, catechin, p-hydroxybenzoic acid, chlorogenic acid, caffeic acid, epicatechin, syringic acid, vanillin, p-coumaric acid, o-coumaric acid, ferulic acid, sinapic acid, benzoic acid, rutin, rosmarinic acid, cinnamic acid, quercetin, kaempferol and luteolin	(Kotiloğlu et al., 2020)
<i>Prosthechea karwinskii</i>	pseudobulbs, leaves and flowers	40% sucrose induced MS male Wistar rats	reducing glucose level, reducing cholesterol and triglycerides levels		flavonoid	(Rojas-Olivos et al., 2017)
<i>Dendrobium officinale</i>	stem	type 2 diabetic rats	ameliorate the symptoms of oxidative stress, inflammation, and hepatic lipid accumulation of liver	metformin	polysaccharide	(Yang et al., 2020)
<i>Dactylorhiza hatagirea</i> (D. hatagirea)	hydroalcoholic extract of root	alloxan monohydrate induced wistar albino rats.	reducing blood glucose, lipid parameters and significantly increasing body weight.	glibenclamide	flavonoids (quercetin), carbohydrate and saponins	(Choukary et al., 2019)
<i>Orchis anatolica</i>	roots ethanol extraction	Alloxan induced diabetic rat	antihyperglycemic effect, correct some biochemical markers induced by diabetes	glibenclamide	flavonoids, triallate, theobromide, and tannins	(Khouri & Daradka, 2013)
<i>Dendrobium gibsonii</i>	whole-plant	α -glucosidase inhibition assay	α -glucosidase inhibitory activity as a noncompetitive inhibitor of α -glucosidase	acarbose	dihydrodengibsinin, dendrogibsol, ephemeranthol A, dengibsinin, nobilone, aloifol I, lusianthridin, denchrysan A and 4-methoxy-9H-fluorene-2,5,9-triol	(Thant et al., 2020)
<i>Dendrobium formosum</i>	whole plant	rat L6 myoblasts	Compounds 1 and 12 showed higher α -glucosidase inhibitory activity. Lusianthridin (6) and	acarbose and orlistat	confusarin (1), hircinol (2), erianthridin (3),	(Inthongkawe et al., 2017)

Orchid	Part of the plant	Model description	Major finding	Control	Isolated Bioactive compound	Reference
Roxb. ex Lindl			moscatilin (11) had higher activity than insulin.		gigantol (4), nudol (5), lusianthridin (6), coelonin (7), dihydroconiferyl dihydro-p-coumarate (8), batatasin III (9), 2,5,7-trihydroxy-4-methoxy-9,10-dihydrophenanthrene (10), moscatilin (11), and 5-methoxy-7-hydroxy-9,10-dihydro-1,4-phenanthrenequinone (12)	
<i>Anacamptis pyramidalis</i> (L.) Rich		The antioxidant potential was evaluated by DPPH, ABTS, CUPRAC and FRAP assays. Enzymes inhibition activity against ACh, BChE, tyrosinase, α -glucosidase, and α -amylase			Disaccharide, Citric acid, Parishin G, Roseoside, Gastrodin derivative, Parishin B, Parishin C, Dihydroxybenzoic acid derivative, Caffeic acid derivative, Acacetin derivative, Oxodihydroxyoctadecenoic acid, Trihydroxyoctadecenoic acid	(Mahomody et al., 2020)
<i>Prosthechea michuacana</i> (Lex.) WE Higgins	bulbs Hexane, chloroform and methanol extracts of <i>P. michuacana</i>	STZ induced diabetic Wistar albino rats	anti-hyperglycemic and antihyperlipemic activity, improving the hyperinsulinemia and produces a significant change on AGEs formation	AGEs formation in vitro with aminoguanidine, antihyperglycemia in vivo with glibenclamide and tolbutamide		(Gutierrez & Hoyos-Vadillo, 2011)
<i>Bauhinia variegata</i>	Young B. variegata L. (var. Candida) leaves	STZ induced diabetic rats	antidiabetic effect through restoring the normal architecture of pancreatic β -cells in addition to the antioxidant and hypolipidemic effect		Polyphenol	(Abdel-Halim et al., 2020)
<i>Gastrodia elata</i> Blume (GE)	GE, fermented by <i>Saccharomyces cerevisiae</i>	High Glucose induced human umbilical vein endothelial cells (HUVECs)	protect against the oxidative stress, and inflammatory conditions in endothelial cells, caused by HG		Phenolic compounds (gastrodin, p-hydroxybenzyl alcohol (HBA), p-	(Kwon et al., 2012)

Orchid	Part of the plant	Model description	Major finding	Control	Isolated Bioactive compound	Reference
<i>Maxillaria tenuifolia</i>	EtOAc extract of the flower	Anti-oxidant activity were determined in ferric thiocyanate method. α -glucosidase inhibitory activity compared with synthetic inhibitor	suppressing carbohydrate disintegration and could prevent damage to organisms by oxidative stress	acarbose	hydroxybenzaldehyde (HBZ)) 3,4-dihydroxy benzoic acid methyl ester (1), flavanthridin (2), vanillic acid (3) and mangiferin (4)	(C. Li, 2021)
<i>Diaphananth e bidens (D. bidens)</i> (AFZEL. EX SW) SCHLTR	methanol leaf extract	STZ induced hyperglycemic rats	reducing blood glucose	tolbutamide	saponins, steroids, tannins and terpenoids	(Ottah et al., 2012)
<i>Bletilla striata</i>	polysaccharides of <i>B. striata</i>	high fat diet (HFD)-fed mice	reducing obesity and metabolic disorders in HFD-fed mice		polysaccharides of <i>B. striata</i>	(Hu et al., 2020)
<i>Dendrobium ofcinale</i>	leaves hot water extraction, alcohol sedimentation and chromatographic separation	inflammatory cell model by LPS acting THP-1 cells	protecting THP-1 cells from LPS-stimulated cytotoxicity, inhibiting reactive oxygen species formation, suppressed toll-like receptor-4 (TLR-4), myeloid differentiation factor (MyD88) and tumour necrosis factor receptor-associated factor-6 (TRAF-6) mRNA and protein expression		polysaccharides	(Zhang et al., 2018)
<i>Dendrobium candidum</i>	dry dendrobium	human corneal epithelial cells (HCEC)	improving the proliferative activity of HCEC cells under the high glucose environment and reduce the apoptosis of cells by regulating the expression of bax and bcl-2. Protecting and repairing corneal epithelial cells damage in high glucose.		polysaccharide	(Q. Li et al., 2017)
<i>Aerides multiflora</i>	dried powder from the whole plants macerated with MeOH	The liberation of p-nitrophenol from the substrate p-nitrophenol- α -D-glucopyranoside (PNPG).	compound revealed a non-competitive inhibition and suggested as a candidate structure for α -glucosidase inhibitor		Aerimultin, Dihydrosinapyl dihydroferulate, 6-Methoxy coelonin, Gigantol, Imbricatin, Agrostonin, Dihydroconiferyl dihydro-p-coumarate, 5-Methoxy-9,10-dihydrophenanthrene-2,3,7-triol, Acarbose	(Thant et al., 2021)

Orchid	Part of the plant	Model description	Major finding	Control	Isolated Bioactive compound	Reference
<i>Eulophia ochreatea</i> L	Tubers extracted with methanol and distilled water	antioxidant activity using DPPH radical scavenging activity, total antioxidant capacity using phosphomolybdenum method, amylase inhibition assay using the chromogenic DNSA method, antiglycation activity using BSA-fructose assay, Nitrobluetetrazolium assay was used to determine the fructosamine	antioxidant activity and favorable α amylase inhibitory activity	ascorbic acid	Phenolic and flavonoids compounds	(Jagtap et al., 2012)
<i>Gymnadenia orchidis</i> Lindl	fibrous root	STZ induced adult female albino mice of BALB C	inhibiting glycation of the hemoglobin, normalizing the lipid profile of diabetic animals, improving antioxidant status and reducing lipid peroxidation, recovering DNA strand breakage		Terpenoids	(Arzoo et al., 2018)
<i>Grammatophyllum speciosum</i> Blume	ethanol extracts prepared from leaf, rhizome+root and pseudobulbs	antioxidant activity using DPPH assay, α -glucosidase inhibition assay was undertaken according to a modified Lebowitz (1998).	DPPH free radical scavenging activity and α -Glucosidase inhibitory activity	Trolox (6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid) and Acarbose		(Rungruchkanont & Chatsuwann, 2019)
<i>Dendrobium christyanum</i> Rchb.f	A methanolic extract from the dried root	in vitro α -glucosidase inhibitory and glucose uptake stimulatory activities	Compounds 4 and 6 appear to be potential hypoglycemic agents since they possess both α -glucosidase inhibitory and glucose uptake stimulatory activities.	acarbose	Methyl haematommate (1), methyl 2,4-dihydroxy-3,6-dimethylbenzoate (3), ndocosyl 4-hydroxy-trans-cinnamate (4), vanillin (5), coniferyl aldehyde (6), 4,5-dihydroxy-2-methoxy-9,10-dihydrophenanthrene (7), gigantol (10), and diorcinolic acid (13).	(San et al., 2020)
<i>Dactylorhiza hatagirea</i>	methanolic leaf extract	α -amylase and α -glucosidase inhibition	inhibiting α -Amylase and α -Glucosidase enzymes, elevating relative expression	Acarbose and Metformin		(Alsawalha et al., 2019)

Orchid	Part of the plant	Model description	Major finding	Control	Isolated Bioactive compound	Reference
		assays and in vitro cellular assays such as glucose uptake assay and glucose transporter type 4 (GLUT4) expression studies in 3T3-L1 cell line	rates of GLUT4 receptor and amount of 2-(N-(7-Nitrobenz-2-oxa-1,3-diazol-4-yl) Amino)-2-Deoxyglucose taken up by 3T3-L1 cells			
<i>Calanthe fimbriata</i> Franch.	roots of <i>C. fimbriata</i> methanol extract	oral glucose tolerance test (OGTT) mice and in streptozotocin (STZ)-induced diabetic mice	decreasing ALT, AST and TG levels, improving hepatomegaly, and increasing hepatic glycogen content	Gliclazide and metformin	more than 40 constituents were determined including organic acids, ester, and sterols, etc.	(Peng et al., 2019)
<i>Dendrobium delacourii</i>	dried powder of whole plant macerated with MeOH and partitioned with ethyl acetate (EtOAc)	α -glucosidase enzyme assay using p-nitrophenyl- α -D-glucoside (PNPG) and anti-adipogenic activity using Mouse embryonic preadipocyte 3T3-L1 cell	α -glucosidase inhibitory activity and anti-adipogenic properties	oxyresveratrol	Phoyunnanin E, phoyunnanin C, densifloral B	(Thant et al., 2022)

Antiglycation Potential

Four orchids of this study, *Gymnadenia orchidis* Lindl, *Eulophia ochreatea* L, *Dendrobium aqueum* Lindl, and *Prosthechea michuacana* (Lex.) WE Higgins have antiglycation activity. Protein glycation is a biomarker for diagnosis of diabetes. Glucose is a directed glycation agent of amino group in proteins, which modifies N-terminal and sidechain amino groups. Afterwards, at later stage, AGEs, are formed by the degradation of fructosamines and by the reaction of reactive dicarbonyl metabolites, such as methylglyoxal (MGO) (Rabbani & Thornalley, 2021). Inhibition of MGO formation is known as one of the antiglycation mechanisms to inhibit AGE formation. In addition to inhibiting its formation, the antiglycation mechanism can also be carried out by acting as a scavenger for this molecule (Jung et al., 2019). Protein glycation and oxidative stress caused by chronic hyperglycemia are the major factors in diabetic complication. Some substances prevent the formation of AGEs by acting as an antioxidant, inhibiting sugar autooxidation, binding to amino groups, early degradation of Maillard Reaction Products (MRP), and by reducing sugars, so that these sugars cannot bind to the amino groups of a protein (Yulianti et al., 2021).

Another mechanism of action of antiglycation compounds is by binding to their receptors, this can cause cellular mechanisms due to the interaction of AGEs with their receptors to be inhibited (Sindhuja et al., 2021). Orchids also contain polyphenols, such as *Bauhinia variegata* (Abdel-Halim et al., 2020). Polyphenols are one of the antiglycation compounds that increase the work of peroxisome proliferator-activated receptors (PPAR), which controls carbohydrate and lipid metabolism, and has been shown to reduce RAGE expression (Di Sotto et al., 2019).

Stimulating Insulin, Anti-insulin Resistance and Elevating GLUT4 Receptor

Insulin is a peptide hormone that binds plasma membrane-bound receptors in target cells. This hormone promotes glucose utilization and storage by increasing glucose transport and glycogen synthesis in skeletal muscle, activating glycogen synthesis, increasing lipogenesis and decreasing gluconeogenic gene expression in liver, as well as suppressing lipolysis and increasing glucose transport and lipogenesis in white adipocyte tissue (Petersen & Shulman, 2018). There are some orchids from this study that improve the

insulin signaling namely *Dactylorhiza hatagirea*, *Prosthechea michuacana* (Lex.) W.E. Higgins and *Prosthechea karwinskii*. Targets of these compounds that affect insulin signaling include insulin receptor substrate, phosphatidylinositol 3-kinase, glucose transporter, activated protein kinase (AMPK), glycogen synthase kinase 3, MAPKs, JNK, NF- κ B, protein tyrosine phosphatase 1B, nuclear factor-E2-related factor 2, and peroxisome proliferator-activated receptors (J. Li et al., 2019). Maintenance and enhancement of β -cell function has the potential to improve diabetes. Specific growth factors, cell cycle mediators, and nuclear factors have been proposed to regulate β -cell homeostasis (Chang et al., 2013).

Anti-Adipogenic Properties

Lipid metabolism, such as increased lipogenesis and decreased lipolysis in adipose tissue occurs largely as a response of glucose homeostasis to insulin stimulation (M. Li et al., 2022). Orchids *Prosthechea karwinskii*, *Prosthechea michuacana* (Lex.) W.E. Higgins, *Dendrobium officinale*, *Dactylorhiza hatagirea*, *Prosthechea michuacana* (Lex.) W.E. Higgins, *Bauhinia variegata*, *Bletilla striata*, *Gymnadenia orchidis* Lindl, *Calanthe fimbriata* Franch, *Dendrobium delacourii* have the properties to improve lipid metabolism in diabetes. Polyphenols, besides functioning as antioxidants also have antiadipogenic abilities. Reduction of adipose tissue is carried out through the mechanism of inhibiting cell proliferation, increasing cell apoptosis, inhibiting differentiation from pre-adipose to adipose, inhibiting cellular lipid accumulation and increasing lipolysis (Nam et al., 2019).

Inhibiting α -Amylase

α -Amylase catalyzes the hydrolysis of starch which will produce glucose as the final product. The catalytic activity of this enzyme can be controlled to reduce glucose production at the postprandial stage, which could be of therapeutic benefit to diabetics (Khadayat et al., 2020). Some α -Amylase inhibitor show their mechanisms by interacting with the key active site residues through an array of hydrophobic interactions and hydrogen bonds (Ogunyemi et al., 2022). *Malaxis rheedei* SW, *Anacamptis pyramidalis* (L.) Rich, *Eulophia ochreate* L, *Dactylorhiza hatagirea* are some orchids with antidiabetic property as an α -Amylase inhibitor. Based on table 1, the orchid *Dactylorhiza romana* subsp. *Georgica* (Kotiloğlu et al., 2020) and *Dactylorhiza hatagirea* (Choukarya et al., 2019) contain quercetin compounds. Quercetin is a compound that has been shown to act as an inhibitor for α amylase. The inhibition mechanism carried out by this compound is by forming molecular interactions with the enzyme binding sites (Oso & Olaoye, 2020).

Inhibiting α -Glucosidase

Glucosidases are required for starch digestion. Hence, α -glucosidase inhibitor is one of the methods for treating diabetes by *suppressing* the digestion of carbohydrates, thus slowing down the process of glucose assimilation which will lead to a significant reduction of plasma glucose and postprandial insulin levels (Choudhury et al., 2018). Some orchids in this study which have this property are *Malaxis rheedei* SW, *Dendrobium gibsonii*, *Dendrobium formosum* Roxb. ex Lindl, *Anacamptis pyramidalis* (L.) Rich, *Maxillaria tenuifolia*, *Aerides multiflora*, *Grammatophyllum speciosum* Blume, *Dendrobium christyanum* Rehb.f, *Dactylorhiza hatagirea*, *Dendrobium delacourii*. Previous study has showed competitive mode of inhibition mechanism of α -Glucosidase inhibitor from some plant substances (Lianza et al., 2022).

Antiinflammation

Metabolic inflammation is involved in diabetes and its complications (such as diabetic nephropathy, retinopathy and neuropathy). Systemic inflammation occurs as a result of an increase in proinflammatory cytokines (IL-6, IL-1 β , TNF- α), which are elicited by chemokines. Increased recruitment of inflammatory cells in metabolic networks, and activation of inflammatory responses occur due to NF- κ B activation and signaling of AMPK and PPAR- γ (Kong et al., 2021). *Prosthechea karwinskii*, *Dendrobium officinale*, *Gastrodia elata* Blume have the antiinflammation activity. Bioactive compounds from plants that can regulate AKT, mTOR, adenosine monophosphate, AMPK, AGEs, growth factors, proinflammatory cytokines (IL-1 β and TNF- α), oxygen species, and various other signaling mechanisms related to diabetes can be projected as an effective therapy (Kaabi, 2022).

Conclusion

In conclusion, orchid has been widely used in traditional medicine. All parts of these plants can be extracted and contain some active substances. Several studies have reported some antidiabetic mechanisms of several species of orchids. These anti-diabetic mechanisms include being antioxidants, anti-glycation and anti-inflammation, increasing insulin action, influencing lipid metabolism, and inhibiting α -amylase and α -glucosidase.

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Author Contributions

EY, ISM and LS constructed the idea for this manuscript and planned the methods. EY, ISM and LS had responsibility in extracting the data from the journals, data management and reporting. EY and TCH analyzed and interpreted the extracting datas. EY had responsibility in the construction of the manuscript. All authors read and approved the final manuscript.

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Conflicts of Interest

There is no conflict of interest.

References

- Abdel-Halim, A. H., Fyiad, A. A. A., Aboulthana, W. M., El-Sammad, N. M., Youssef, A. M., & Ali, M. M. (2020). Assessment of the anti-diabetic effect of Bauhinia variegata gold nano-extract against streptozotocin induced diabetes mellitus in rats. *Journal of Applied Pharmaceutical Science*, 10(5), 77–91. <https://doi.org/10.7324/JAPS.2020.10511>
- Ahmed, O. A. A., Azhar, A. S., Tarkhan, M. M., Balamash, K. S., & El-Bassossy, H. M. (2020). Antglycation Activities and Common Mechanisms Mediating Vasculoprotective Effect of Quercetin and Chrysin in Metabolic Syndrome. *Evidence-Based Complementary and Alternative Medicine*, 2020. <https://doi.org/10.1155/2020/3439624>
- Alsawalha, M., Al-Subaei, A., Al-Jindan, R., Bolla, S., Sen, D., Balakrishna, J., Ravi, P., Reddy Gollapalli, S., Veeraraghavan, V., Pillai, A., Joseph, J., Salahuddin, M., & Mohan, S. (2019). Anti-diabetic activities of Dactylorhiza hatagirea leaf extract in 3T3-L1 cell line model. *Pharmacognosy Magazine*, 15(64), 212. https://doi.org/10.4103/pm.pm_8_19
- Arzoo, S. H., Chattopadhyay, K., Banerjee, S., & Chattopadhyay, B. (2018). Synergistic improved efficacy of Gymnadenia orchidis root Salep and pumpkin seed on induced diabetic complications. *Diabetes Research and Clinical Practice*, 146, 278–288. <https://doi.org/10.1016/j.diabres.2018.10.025>
- Ashu Rajeshbhai, V., & Ingalthalli, R. (2022). A Review Article on: An Overview of Medicinal Importance of Orchids. *Acta Scientific Medical Sciences*, 6(7), 28–36.
- Barragán-Zarate, G. S., Alexander-Aguilera, A., Lagunez-Rivera, L., Solano, R., & Soto-Rodríguez, I. (2021). Bioactive compounds from Prosthechea karwinskii decrease obesity, insulin resistance, pro-inflammatory status, and cardiovascular risk in Wistar rats with metabolic syndrome. *Journal of Ethnopharmacology*, 279. <https://doi.org/10.1016/j.jep.2021.114376>
- Barragán-Zarate, G. S., Lagunez-Rivera, L., Solano, R., Pineda-Peña, E. A., Landa-Juárez, A. Y., Chávez-Piña, A. E., Carranza-Álvarez, C., & Hernández-Benavides, D. M. (2020). Prosthechea karwinskii, an orchid used as traditional medicine, exerts anti-inflammatory activity and inhibits ROS. *Journal of Ethnopharmacology*, 253. <https://doi.org/10.1016/j.jep.2020.112632>
- Chang, C. L. T., Lin, Y., Bartolome, A. P., Chen, Y. C., Chiu, S. C., & Yang, W. C. (2013). Herbal therapies for type 2 diabetes mellitus: Chemistry, biology, and potential application of selected plants and compounds. In *Evidence-based Complementary and Alternative Medicine* (Vol. 2013). <https://doi.org/10.1155/2013/378657>
- Choudhury, H., Pandey, M., Hua, C. K., Mun, C. S., Jing, J. K., Kong, L., Ern, L. Y., Ashraf, N. A., Kit, S. W., Yee, T. S., Pichika, M. R., Gorain, B., & Kesharwani, P. (2018). An update on natural compounds in the remedy of diabetes mellitus: A systematic review. *Journal of Traditional and Complementary Medicine*, 8(3), 361–376. <https://doi.org/10.1016/J.JTCME.2017.08.012>
- Choukarya, R., Choursia, A., & Rath, J. (2019). In Vivo and In Vitro Antidiabetic Activity of Hydroalcoholic Extract of Dactylorhiza Hatagirea Roots: An Evaluation of Possible Phytoconstituents. *Journal of Drug Delivery and Therapeutics*, 9(6-s), 76–81. <https://doi.org/10.22270/jddt.v9i6-s.3752>
- Di Sotto, A., Locatelli, M., Macone, A., Toniolo, C., Cesa, S., Carradori, S., Eufemi, M., Mazzanti, G., & Di Giacomo, S. (2019). Hypoglycemic, antiglycation, and cytoprotective properties of a phenol-rich extract from waste peel of punica granatum L. Var. Dente di cavallo DC2. *Molecules*, 24(17). <https://doi.org/10.3390/molecules24173103>
- Francenia Santos-Sánchez, N., Salas-Coronado, R., Villanueva-Cañongo, C., & Hernández-Carlos, B. (2019). Antioxidant Compounds and Their Antioxidant Mechanism. In *Antioxidants*. IntechOpen. <https://doi.org/10.5772/intechopen.85270>
- Gutierrez, P., & Hoyo-Vadillo, C. (2011). Anti-diabetic activity of an hexane extract of Prosthechea michuacanain in streptozotocin-induced diabetic rats. *Boletín Latinoamericano y Del Caribe de Plantas Medicinales y Aromáticas*, 10(6), 570–580. Retrieved from www.blacpma.usach.cl
- Gutierrez, R. M. P. (2013). Evaluation of the Hypoglycemic and Hypolipidemic Effects of Triterpenoids from Prosthechea michuacana in Streptozotocin-induced Type 2 Diabetic Mice.

- Pharmacologia*, 4, 170-179. Retrieved from <https://scialert.net/fulltext/?doi=pharmacologia.2013.170.179>
- Haridas, R., P, S., & Thekkan, S. (2017). In-vitro antidiabetic activity of *Malaxis rheedei* SW (whole plant): an endangered medicinal orchid. *International Journal of Pharma and Bio Sciences*, 8(2). <https://doi.org/10.22376/ijpbs.2017.8.2.p130-133>
- Hu, B., Ye, C., Leung, E. L. H., Zhu, L., Hu, H., Zhang, Z., Zheng, J., & Liu, H. (2020). *Bletilla striata* oligosaccharides improve metabolic syndrome through modulation of gut microbiota and intestinal metabolites in high fat diet-fed mice. *Pharmacological Research*, 159. <https://doi.org/10.1016/j.phrs.2020.104942>
- Hunyadi, A. (2019). The mechanism(s) of action of antioxidants: From scavenging reactive oxygen/nitrogen species to redox signaling and the generation of bioactive secondary metabolites. In *Medicinal Research Reviews* (Vol. 39, Issue 6, pp. 2505-2533). John Wiley and Sons Inc. <https://doi.org/10.1002/med.21592>
- Inthongkaew, P., Chatsumpun, N., Supasuteekul, C., Kitisripanya, T., Putalun, W., Likhitwitayawuid, K., & Sritularak, B. (2017). α -glucosidase and pancreatic lipase inhibitory activities and glucose uptake stimulatory effect of phenolic compounds from *dendrobium formosum*. *Revista Brasileira de Farmacognosia*, 27(4), 480-487. <https://doi.org/10.1016/j.bjpr.2017.05.005>
- Jagtap, S., Narkhede, A., Nirmal, P., Tupe, R., Kulkarni, O., & Harsulkar, A. (2012). In vitro Antioxidant, Antiglycation and α -amylase inhibitory potential of *Eulophia ochreatea* L. *Journal of Pharmacy Research*, 5(5), 2532-2537. Retrieved from www.jpronline.info
- Ji, M., Gong, X., Li, X., Wang, C., & Li, M. (2020). Advanced research on the antioxidant activity and mechanism of polyphenols from *hippophae* species-a review. In *Molecules* (Vol. 25, Issue 4). MDPI <https://doi.org/10.3390/molecules25040917>
- Joshi, P. R., Paudel, M. R., Chand, M. B., Pradhan, S., Pant, K. K., Joshi, G. P., Bohara, M., Wagner, S. H., Pant, B., & Pant, B. (2020). Cytotoxic effect of selected wild orchids on two different human cancer cell lines. *Heliyon*, 6(5). <https://doi.org/10.1016/j.heliyon.2020.e03991>
- Jung, E., Park, S. Bin, Jung, W. K., Kim, H. R., & Kim, J. (2019). Antiglycation activity of aucubin in vitro and in exogenous methylglyoxal injected rats. *Molecules*, 24(20). <https://doi.org/10.3390/molecules24203653>
- Kaabi, Y. A. (2022). Potential Roles of Anti-Inflammatory Plant-Derived Bioactive Compounds Targeting Inflammation in Microvascular Complications of Diabetes. In *Molecules* (Vol. 27, Issue 21). MDPI. <https://doi.org/10.3390/molecules27217352>
- Khadayat, K., Marasini, B. P., Gautam, H., Ghaju, S., & Parajuli, N. (2020). Evaluation of the alpha-amylase inhibitory activity of Nepalese medicinal plants used in the treatment of diabetes mellitus. *Clinical Phytoscience*, 6(1). <https://doi.org/10.1186/s40816-020-00179-8>
- Khouri, N. A., & Daradka, H. (2013). Antidiabetic effect of *Orchis anatolica* root extracts on alloxan-induced diabetic rats. *Comparative Clinical Pathology*, 22(3), 347-354. <https://doi.org/10.1007/s00580-012-1415-8>
- Kong, M., Xie, K., Lv, M., Li, J., Yao, J., Yan, K., Wu, X., Xu, Y., & Ye, D. (2021). Anti-inflammatory phytochemicals for the treatment of diabetes and its complications: Lessons learned and future promise. *Biomedicine & Pharmacotherapy*, 133, 110975. <https://doi.org/10.1016/J.BIOPHA.2020.110975>
- Kooti, W., Farokhipour, M., Asadzadeh, Z., Ashtary-Larky, D., & Asadi-Samani, M. (2016). The role of medicinal plants in the treatment of diabetes: a systematic review. *Electronic Physician*, 8(1), 1832-1842. <https://doi.org/10.19082/1832>
- Kotiloglu, D., Acet, T., & Özcan, K. (2020). Phytochemical profile and biological activity of a therapeutic orchid from Anatolia: *Dactylorhiza romana* subsp. *georgica*. *Journal of Food Measurement and Characterization*, 14(6), 3310-3318. <https://doi.org/10.1007/s11694-020-00566-2>
- Kuo, Y. J., Pei, J. K., & Chao, W. W. (2022). Pharmacological and Chemical Potential of *Spiranthes sinensis* (Orchidaceae): A Narrative Review. In *Plants* (Vol. 11, Issue 13). MDPI. <https://doi.org/10.3390/plants11131692>
- Kwon, S.-U., Jeon, S.-B., Xin, M., Kim, J.-H., Im, J.-Y., Cha, J.-Y., Jee, H.-K., Lee, O.-G., Kim, D.-K., & Lee, Y.-M. (2012). Inhibitory Effects of Fermented *Gastrodia elata* on High Glucose-induced NO and IL-8 Production in Human Umbilical Vein Endothelial Cells. In *Natural Product Sciences* (Vol. 18, Issue 4). Retrieved from <https://www.dbpia.co.kr/Journal/articleDetail?nodeId=NODE11129699>
- Li, C. (2021). Constituents of the Flower of *Maxillaria tenuifolia* and Their Anti-Diabetic Activity. *Records of Natural Products*, 1, 1-6. <https://doi.org/10.25135/rnp.274.2106.2093>
- Li, J., Bai, L., Wei, F., Zhao, J., Wang, D., Xiao, Y., Yan, W., & Wei, J. (2019). Therapeutic mechanisms of herbal medicines against insulin resistance: A

- review. In *Frontiers in Pharmacology* (Vol. 10, Issue JUN). Frontiers Media S.A. <https://doi.org/10.3389/fphar.2019.00661>
- Li, M., Chi, X., Wang, Y., Setrerrahmane, S., Xie, W., & Xu, H. (2022). Trends in insulin resistance: insights into mechanisms and therapeutic strategy. In *Signal Transduction and Targeted Therapy* (Vol. 7, Issue 1). Springer Nature. <https://doi.org/10.1038/s41392-022-01073-0>
- Li, Q., Chen, J., Li, Y., Chen, T., Zou, J., & Wang, H. (2017). Effect of polysaccharide of dendrobium candidum on proliferation and apoptosis of human corneal epithelial cells in high glucose. *Medicine (United States)*, 96(32). <https://doi.org/10.1097/MD.00000000000007773>
- Lianza, M., Poli, F., Nascimento, A. M. do, Soares da Silva, A., da Fonseca, T. S., Toledo, M. V., Simas, R. C., Chaves, A. R., Leitão, G. G., & Leitão, S. G. (2022). In vitro α -glucosidase inhibition by Brazilian medicinal plant extracts characterised by ultra-high performance liquid chromatography coupled to mass spectrometry. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 37(1), 554–562. <https://doi.org/10.1080/14756366.2021.2022658>
- Mahomoodally, M. F., Marie Carene, M. C. N., Zengin, G., Eulogio, E. J., Abdullah, H. H., Ak, G., Senkardes, I., Chiavaroli, A., Menghini, L., Recinella, L., Brunetti, L., Leone, S., Orlando, G., & Ferrante, C. (2020). Phytochemical analysis, network pharmacology and in silico investigations on anacamptis pyramidalis tuber extracts. *Molecules*, 25(10). <https://doi.org/10.3390/molecules25102422>
- Martha, E., & Rahayu, D. (2022). Review of the conservation efforts of orchid species in Bogor Botanic Gardens. Orchid Phytogeography View project. *Proceedings of the 2021 Virtual World Orchid Conference*. Retrieved from <https://www.researchgate.net/publication/359104195>
- Martha Pérez Gutiérrez, R. (2010). Orchids: A review of uses in traditional medicine, its phytochemistry and pharmacology. *Journal of Medicinal Plants Research*, 4(8), 592–638. <https://doi.org/10.5897/JMPR10.012>
- Minh, T. N., Khang, D. T., Tuyen, P. T., Minh, L. T., Anh, L. H., Van Quan, N., Ha, P. T. T., Quan, N. T., Toan, N. P., Elzaawely, A. A., & Xuan, T. D. (2016). Phenolic compounds and antioxidant activity of phalaenopsis orchid hybrids. *Antioxidants*, 5(3). <https://doi.org/10.3390/antiox5030031>
- Mukherjee, S., Jagtap, S., Vidyapeeth, B., & Tupe, R. (2012). Antiglycation and antioxidant activity of a rare medicinal orchid *Dendrobium aequum* Lindl. In *Medicinal Chemistry & Drug Discovery* (Vol. 2012, Issue 2). Retrieved from <https://www.researchgate.net/publication/235751078>
- Musharof Hossain, M. (2011). Therapeutic orchids: traditional uses and recent advances—An overview. *Fitoterapia*, 82(2), 102–140. <https://doi.org/10.1016/J.FITOTE.2010.09.007>
- Nam, W., Nam, S. H., Kim, S. P., Levin, C., & Friedman, M. (2019). Anti-adipogenic and anti-obesity activities of purpurin in 3T3-L1 preadipocyte cells and in mice fed a high-fat diet. *BMC Complementary and Alternative Medicine*, 19(1). <https://doi.org/10.1186/s12906-019-2756-5>
- Nawaz, H., Shad, M. A., Rehman, N., Andaleeb, H., & Ullah, N. (2020). Effect of solvent polarity on extraction yield and antioxidant properties of phytochemicals from bean (*Phaseolus vulgaris*) seeds. *Brazilian Journal of Pharmaceutical Sciences*, 56. <https://doi.org/10.1590/s2175-97902019000417129>
- Nimse, S. B., & Pal, D. (2015). Free radicals, natural antioxidants, and their reaction mechanisms. *RSC Advances*, 5(35), 27986–28006. <https://doi.org/10.1039/c4ra13315c>
- Ogunyemi, O. M., Gyebe, G. A., Saheed, A., Paul, J., Nwaneri-Chidozie, V., Olorundare, O., Adebayo, J., Koketsu, M., Aljarba, N., Alkahtani, S., Batiha, G. E. S., & Olaiya, C. O. (2022). Inhibition mechanism of alpha-amylase, a diabetes target, by a steroidal pregnane and pregnane glycosides derived from *Gongronema latifolium* Benth. *Frontiers in Molecular Biosciences*, 9. <https://doi.org/10.3389/fmolb.2022.866719>
- Oso, B. J., & Olaoye, I. F. (2020). Antiglycaemic potentials and molecular docking studies of the extracts of *Cassia alata* L. *Beni-Suef University Journal of Basic and Applied Sciences*, 9(1). <https://doi.org/10.1186/s43088-020-00068-6>
- Ottah, A. A., Augustine, O., Obiora, I. O., & Maxwell, E. (2012). Antihyperglycemic effects of the methanol leaf extract of *Diaphanthe bidens* in normoglycemic and streptozotocin-induced hyperglycemic rats Asian Pacific Journal of Tropical Medicine *Diaphanthe bidens* Streptozotocin Tolbutamide Normoglycaemic Hyperglycemic. In *Asian Pacific Journal of Tropical Medicine*. Retrieved from www.elsevier.com/locate/apjtm
- Pan, L. H., Li, X. F., Wang, M. N., Zha, X. Q., Yang, X. F., Liu, Z. J., Luo, Y. B., & Luo, J. P. (2014). Comparison of hypoglycemic and antioxidative effects of polysaccharides from four different dendrobium species. *International Journal of Biological*

- Macromolecules*, 64, 420-427. <https://doi.org/10.1016/j.ijbiomac.2013.12.024>
- Paudel, M. R., Chand, M. B., Pant, B., & Pant, B. (2019). Assessment of antioxidant and cytotoxic activities of extracts of *Dendrobium crepidatum*. *Biomolecules*, 9(9). <https://doi.org/10.3390/biom9090478>
- Peng, Y., Gao, Y., Zhang, X., Zhang, C., Wang, X., Zhang, H., Wang, Z., Liu, Y., & Zhang, H. (2019). Antidiabetic and hepatoprotective activity of the roots of *Calanthe fimbriata* Franch. *Biomedicine and Pharmacotherapy*, 111, 60-67. <https://doi.org/10.1016/j.biopha.2018.12.066>
- Petersen, M. C., & Shulman, G. I. (2018). Mechanisms of Insulin Action and Insulin Resistance. *Physiol Rev*, 98, 2133-2223. <https://doi.org/10.1152/physrev>
- Rabbani, N., & Thornalley, P. J. (2021). Protein glycation - biomarkers of metabolic dysfunction and early-stage decline in health in the era of precision medicine. *Redox Biology*, 42. <https://doi.org/10.1016/j.redox.2021.101920>
- Rojas-Olivos, A., Solano-Gómez, R., Alexander-Aguilera, A., Jiménez-Estrada, M., Zilli-Hernández, S., & Lagunez-Rivera, L. (2017). Effect of *Prosthechea karwinskii* (Orchidaceae) on obesity and dyslipidemia in Wistar rats. *Alexandria Journal of Medicine*, 53(4), 311-315. <https://doi.org/10.1016/j.ajme.2016.11.004>
- Rungrouchkanont, K., & Chatsuwana, Y. (2019). Antioxidant and α -glucosidase inhibitor activities of *Grammatophyllum speciosum* Blume. *Acta Horticulturae*, 1245, 73-78. <https://doi.org/10.17660/ActaHortic.2019.1245.11>
- Safari, M. R., Azizi, O., Heidary, S. S., Kheiripour, N., & Ravan, A. P. (2018). Antiglycation and antioxidant activity of four Iranian medical plant extracts. *Journal of Pharmacopuncture*, 21(2), 82-89. <https://doi.org/10.3831/KPI.2018.21.010>
- Salleh, N. H., Zulkipli, I. N., Mohd Yasin, H., Ja' Afar, F., Ahmad, N., Wan Ahmad, W. A. N., & Ahmad, S. R. (2021). Systematic Review of Medicinal Plants Used for Treatment of Diabetes in Human Clinical Trials: An ASEAN Perspective. In *Evidence-based Complementary and Alternative Medicine* (Vol. 2021). Hindawi Limited. <https://doi.org/10.1155/2021/5570939>
- San, H. T., Boonsongcheep, P., Putalun, W., Mekboonsonglarp, W., Sritularak, B., & Likhitwitayawuid, K. (2020). α -Glucosidase Inhibitory and Glucose Uptake Stimulatory Effects of Phenolic Compounds From *Dendrobium christyanum*. *Natural Product Communications*, 15(3). <https://doi.org/10.1177/1934578X20913453>
- Sindhuja, A., Vimalavathini, R. R., & Kavimani, S. (2021). In Silico Docking Studies of Antiglycation Activity of Isorhamnetin on Molecular Proteins of Advanced Glycation End Product (AGE) Pathway. *Biomedical and Pharmacology Journal*, 14(4), 2299-2306. <https://doi.org/10.13005/bpj/2331>
- Singh, S., Singh, A. K., Kumar, S., Kumar, M., Pandey, P. K., & Singh, M. C. K. (2012). Medicinal properties and uses of orchids: a concise review. *Elixir Appl. Botany*, 52(2012), 11627-11634. Retrieved from <https://www.researchgate.net/publication/292131192>
- Thant, M. T., Chatsumpun, N., Mekboonsonglarp, W., Sritularak, B., & Likhitwitayawuid, K. (2020). New Fluorene Derivatives from *Dendrobium gibsonii* and Their α -Glucosidase Inhibitory Activity. *Molecules (Basel, Switzerland)*, 25(21). <https://doi.org/10.3390/molecules25214931>
- Thant, M. T., Khine, H. E. E., Nealiga, J. Q. L., Chatsumpun, N., Chaotham, C., Sritularak, B., & Likhitwitayawuid, K. (2022). α -Glucosidase Inhibitory Activity and Anti-Adipogenic Effect of Compounds from *Dendrobium delacourii*. *Molecules*, 27(4). <https://doi.org/10.3390/molecules27041156>
- Thant, M. T., Sritularak, B., Chatsumpun, N., Mekboonsonglarp, W., Punpreuk, Y., & Likhitwitayawuid, K. (2021). Three novel biphenanthrene derivatives and a new phenylpropanoid ester from *aerides multiflora* and their α -glucosidase inhibitory activity. *Plants*, 10(2), 1-14. <https://doi.org/10.3390/plants10020385>
- Tsering, J., Tam, N., Tag, H., Gogoi, B. J., & Apang, O. (2017). Medicinal Orchids of Arunachal Pradesh: A Review. *Bulletin of Arunachal Forest Research*, 32(2), 1-16. Retrieved from <https://rb.gy/j5v7w>
- Yang, J., Chen, H., Nie, Q., Huang, X., & Nie, S. (2020). *Dendrobium officinale* polysaccharide ameliorates the liver metabolism disorders of type II diabetic rats. *International Journal of Biological Macromolecules*, 164, 1939-1948. <https://doi.org/10.1016/j.ijbiomac.2020.08.007>
- Yeh, W. J., Hsia, S. M., Lee, W. H., & Wu, C. H. (2017). Polyphenols with antiglycation activity and mechanisms of action: A review of recent findings. *Journal of Food and Drug Analysis*, 25(1), 84-92. <https://doi.org/10.1016/j.jfda.2016.10.017>
- Yulianti, E., Sunarti, & Wahyuningsih, M. S. H. (2021). The effect of *Kappaphycus alvarezii* fraction on plasma glucose, Advanced Glycation End-products formation, and renal RAGE gene expression. *Heliyon*, 7(1), e05978. <https://doi.org/10.1016/j.heliyon.2021.e05978>

- Yulianti, E., Sunarti, & Wahyuningsih, M. S. H. (2022). The effect of *Kappaphycus alvarezii* active fraction on oxidative stress and inflammation in streptozotocin and nicotinamide-induced diabetic rats. *BMC Complementary Medicine and Therapies*, 22(1). <https://doi.org/10.1186/s12906-021-03496-8>
- Zhang, M., Wu, J., Han, J., Shu, H., & Liu, K. (2018). Isolation of polysaccharides from *Dendrobium officinale* leaves and anti-inflammatory activity in LPS-stimulated THP-1 cells. *Chemistry Central Journal*, 12(1). <https://doi.org/10.1186/s13065-018-0480-8>